



Multi-Resistant *Acinetobacter Baumannii* related Septicemia Treatment in Burn Patients using Ampicillin-Sulbactam

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Original article

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Summary

The aim of this study is to evaluate the efficacy of using Ampicillin-Sulbactam (AS) for treatment of septicemia, caused by multi-resistant strains of *Acinetobacter baumannii* (AB), in burn patients. Forty patients were included in the analysis over 2003.

Cultures were performed on blood samples taken from these patients. Sensitivity of the strains to Ampicillin-Sulbactam was determined, marking the zones of inhibited growth and determining the Minimal Inhibitory Concentration (MIC) by using the BIOMIC system.

AB strains in 24/40 patients were shown to be completely resistant to AS.

In 10/40 or 25% of cases the isolate demonstrated medium sensitivity to AS, whereas in only 6/40 or 15% of the patients this strain proved to be completely AS-sensitive.

29/40 Patients were subjected to AS therapy. 16/29 Patients had an AB strain resistant to AS whereas 13/29 Patients showed medium or complete AS sensitivity. These 29 patients were all treated with a maximum daily dose of 3 grams of AS every six hours.

Eradication of AB from the wound bed, after completing the full course, was achieved in 20/29 patients, who fully recovered. This included 9/29 patients that demonstrated in-vitro AS resistance.

In 7/29 patients, despite the completed AS course with a maximum daily dose, a sterile blood culture and eradication of AB from the wound bed was not achieved. These patients died as a consequence.

Key words: Bacteremia, Burns, *Acinetobacter baumannii*, Ampicillin-Sulbactam

INTRODUCTION

Acinetobacter Baumannii (AB) is recognised to contribute significantly to burn patients' morbidity and mortality. AB strains, due to their virulence, may cause septicemia, infection of the lungs, - urinary system, - peritoneum, and wound infection.[1]

The occurrence of infection caused by AB has increased substantially during the last decade. This is

mainly due to the increase in resistance to many antibiotic groups. As a result it has become one of the most common and lethal pathogens to cause hospital infections, including septicemia in burn patients (Table 1).

Multi-resistance of AB comprises beta-lactam antibiotics, aminoglycosides, and chinolones. This causes concern because the potential for treatment of AB infections, including treatment of bacteremia, is diminishing. [2,3]

Table 1. Etiological Factors of Blood Infections Occurring in Burn Patients Hospitalised in Our Centre between 2002 and 2003

No.	Microorganisms	2002		2003	
		Number of strains	Percentage	Number of strains	Percentage
1.	<i>Acinetobacter baumannii</i>	58	28,29%	90	25,5%
2.	<i>Staphylococcus aureus</i> (metacycline-resistant strains)	43	20,97%	77	21,8%
3.	<i>Pseudomonas aeruginosa</i>	35	17,03%	58	16,4%
4.	<i>Staphylococcus epidermidis</i> (metacycline-resistant strains)	25	12,19%	57	16,1%
5.	<i>Staphylococcus aureus</i>	19	9,27%	15	4,2%
6.	<i>Enterococcus faecalis</i>	10	4,88%	10	2,8%
7.	<i>Klebsiella pneumoniae</i>	6	2,93%	12	3,4%
8.	<i>Escherichia coli</i>	5	2,44%	8	2,3%
9.	<i>Candida albicans</i>	4	1,95%	2	0,6%
10.	<i>Staphylococcus epidermidis</i> MSSE	-	-	10	2,8%
11.	<i>Streptococcus pyogenes</i>	-	-	6	1,7%
12.	<i>Proteus mirabilis</i>	-	-	3	0,8%

Table 2. Etiological Factors of Blood Infections Occurring in Burn Patients Who Died (N = 7) between 2002 and 2003

No.	Microorganisms	2002		2003	
		Number of strains	Percentage	Number of strains	Percentage
1.	<i>Acinetobacter baumannii</i>	29	33,3%	43	28,6%
2.	<i>Staphylococcus aureus</i> (metacycline-resistant)	14	16,1%	26	17,4%
3.	<i>Pseudomonas aeruginosa</i>	14	16,1 %	41	27 %
4.	<i>Staphylococcus epidermidis</i> (metacycline-resistant strains)	8	9,1%	16	10,7%
5.	<i>Staphylococcus aureus</i>	8	9,1%	3	2 %
6.	<i>Enterococcus faecalis</i>	5	5,8%	6	4 %
7.	<i>Klebsiella pneumoniae</i>	3	3,5%	7	4,66%
8.	<i>Escherichia coli</i>	5	5,8%	-	-
9.	<i>Candida albicans</i>	1	1,2%	2	1,3 %
10.	<i>Staphylococcus epidermidis</i>	-	-	2	1,3 %

Literature indicates that the use of Ampicillin-Sulbactam (AS) in treating infections caused by multi-resistant strains of AB is clinically effective [4,5,6].

The aim of this study was to evaluate clinical efficacy of the use of AS for infection of the wound bed by multi-resistant strains of AB in burn patients.

The study was carried out in 2003 at the Centre for Burn Treatment in Siemianowice Śląskie, the largest hospital for burn treatment in Poland.

PATIENTS AND METHODS

N = 40 patients with a wound bed infection caused by AB strains were included in the analysis. Patients admitted to the center were selected, using the following diagnostic criteria for of a wound bed infection:

Fever > 39° C, chills, systolic blood pressure < 90mmHg, leukocytosis count > 12.000 and isolation of AB from the wound bed.

Blood cultures were performed by marking sensitivity to AS, using Antimicrobial Disk Susceptibility Tests and marking of MIC with the Biomic system.

RESULTS

In N = 24/40 patients an AB strain was isolated, shown to be completely resistant to AS. In N = 10/40 patients, the isolated strain showed medium sensitivity to AS and in 6/40 patients the strain was sensitive to AS.

Ampicillin-Sulbactam therapy was applied in 29/40 patients of which 16/29 patients had an AB strain com-

pletely resistant to AS. In 13/29 patients there was medium or complete AS-sensitivity. These 29 patients received the maximum daily dose of the above mentioned antibiotic of 3 grams, every 6 hours.

In 20/29 patients, after completing the full course of treatment, eradication of AB from the wound bed was achieved, followed by a complete recovery. This including 9/29 patients with complete resistancy to AS.

In 7/29 patients, despite completing the full AS course, sterile blood cultures and eradication of AB from the wound bed, was not achieved. These patients died as a result of the infection (The overview is given in Table 8).

DISCUSSION

Acinetobacter baumannii strains are mostly resistant to antibiotics from the group of penicillin and cephalosporins. Our observations over a period of the last three years, show resistance of > 80% to cephalosporins of the third and fourth generation.

Resistance to carbapenems within the Acinetobacter baumannii types is increasing. Until recently, carbapenems were among the most effective beta-lactam antibiotics, with little known resistance. [7] Resistance is mainly found in the strains from patients in intensive therapy - and burn wards. This resistance is usually accompanied by resistance to other antibiotic groups. [6]

Even though imipenem is more active than meropenem and is still considered the most effective antibiotic

Table 3. Antibiotic Sensitivity of the Acinetobacter baumannii Strains in 2001 till 2003

Antibiotic	2001r.	2002r.	2003r	Fluctuation of resistance
Colistin	2,5%	1,0%	0,0%	↓
Netilmicin	28,5%	26,1%	38,6%	↓↑
Ampicillin-Sulbactam	32,4%	37,4%	51,5%	↑↑
Meropenem	36,5%	51,7%	67,6%	↑↑
Imipenem	37,2%	52,5%	67,6%	↑↑
Piperacillin-Tazobactam	75,2%	76,9%	93,4%	↑↑
Ceftazidime	88,6%	78,2%	93,8%	↓↑
Amikacin	87,0%	90,2%	97,7%	↑↑
Gentamicin	90,9%	92,5%	97,7%	↑↑
Ciprofloxacin	97,1%	86,3%	94,8%	↓↑
Cefotaxime	97,2%	93,4%	98,5%	↓↑
Cefepime	0,0%	60,4%	86,4%	↑↑

Table 4. Analysis of Patients with Blood Infection Caused by a Strain of Resistant Acinetobacter baumannii Who Were Treated with Ampicillin-Sulbactam

Patients in Whom Eradication of the Microorganism from Blood Was Achieved

No.	Sex	Age	% TBSA	Third-degree burns
1.	M	42	55%	25%
2.	M	41	41%	5%
3.	M	28	62 %	38 %
4.	M	23	51 %	38 %
5.	M	41	49 %	45 %
6.	M	43	47 %	10 %
7.	M	37	56 %	2%
8.	M	42	60 %	40 %
9.	F	60	47 %	10 %
Average value		39	52 %	23 %

against *Acinetobacter* species, however it is not effective for all strains [1]. Sensitivity studies of AB in our Centre in 2001-2003 indicate an ever-increasing resistance to carbapenems, of > 70%.

Lack of sensitivity to aminoglycosides, such as gentamicin and amikacin may be due to various resistance mechanisms, such as the production of enzymes.

We noted that the highest sensitivity (60%-70%) of AB for aminoglycosides was observed to be netilmicin. However, this antibiotic could not be administered, due to insufficient availability to our Center.

Until the beginning of the 1990's, new fluoroquinolones demonstrated effectivity against AB strains; however, in the last decade there has been a rapid growth of strains which are resistant to this group [1]. We noted that 85-95% of the strains were found to be resistant to chinolons.

A combination of beta-lactam antibiotics with beta-lactamase inhibitors, such as sulbactam, may in certain situations, represent an alternative for the treatment of severe infections caused by multi-resistant AB strains [8, 9].

Our observations indicate an in-vitro effectivity for ampicillin with sulbactam to AB strains; the sensitivity approached 50-60 % (Table 3).

Knowledge in the area of epidemiology, microbiology, antibiotic therapy and clinical experience are indispensable in the prevention of infections caused by multi-resistant strains. Limiting of these epidemics is possible only through working with multidisciplinary teams. Standards should be implemented for effective hospital hygiene. The importance of hand – and surface hygiene, as well as education of personnel should be taken on board.

Table 5. Analysis of Patients with Blood Infection Caused by a Strain of Resistant *Acinetobacter baumannii*, Who Were Treated with Ampicillin-Sulbactam

Patients in Whom Eradication of the Microorganism from Blood Was Not Achieved

No.	Sex	Age	% TBSA	Third-degree burns
1.	F	75	43%	28%
2.	F	81	30%	15%
3.	M	48	77%	66 %
4.	M	32	90 %	80 %
5.	M	23	56 %	35 %
6.	F	78	49 %	20 %
7.	M	52	30 %	18%
Average value		55	53,5 %	37 %

Table 6. Analysis of Selected Diagnostic Parameters in Patients with Blood Infection Caused by Resistant *Acinetobacter baumannii*, Who Were Treated with Ampicillin-Sulbactam (MIC 48)

Patients in Whom Eradication of the Microorganism from Blood Was Achieved

No.	Antimicrobial Disk Susceptibility Tests	MIC	Albumin Level	Leukocyte Level	Type of Infection
1	6 mm (R)	48	1,8	19,4	Multi-
2.	9 mm(R)	48	2,6	9,5	Multi-
3.	6 mm(R)	48	2,3	17,9	Multi-
4.	6 mm(R)	48	2,3	7,6	Multi-
5.	10mm(R)	32	2,9	9,1	Multi-
6.	6 mm(R)	48	2,8	10,9	Multi-
7.	11mm(R)	24	2,5	10,7	Single-
8.	10mm(R)	32	2,6	9,4	Single-
9.	10 mm(R)	32	2,1	4,5	Single-
Average value			2,4	11 thousands	

Table 7. Analysis of Selected Diagnostic Parameters in Patients with Blood Infection Caused by a Resistant *Acinetobacter baumannii* Who Were Treated with Ampicillin-Sulbactam (MIC 48)

Patients in Whom Eradication of the Microorganism from Blood Was Not Achieved

No.	Antimicrobial Disk Susceptibility Tests	MIC	Albumin Level	Leukocyte Level	Type of infection
1	6 mm (R)	48	2,2	7,7	Multi-microbial
2.	6 mm(R)	48	1,6	11,9	Single-microbial
3.	6 mm(R)	48	1,2	14,1	Single-microbial
4.	6 mm(R)	48	1,7	12,7	Multi-microbial
5.	6 mm(R)	48	1,7	10,9	Multi-microbial
6.	6 mm(R)	48	1,7	16,2	Multi-microbial
7.	6mm(R)	48	1,9	6,7	Single-microbial
Average value			1,7	11,4 thousands	

Monitoring of hospital infections and an appropriate antibiotic policy are among the most important measures to reduce the incidence of hospital infections caused by multi-resistant pathogens. [1,10,11,13]

Multi-resistant, hospital-acquired infections should be viewed as a primary issue and not just attributed to a specific hospital.

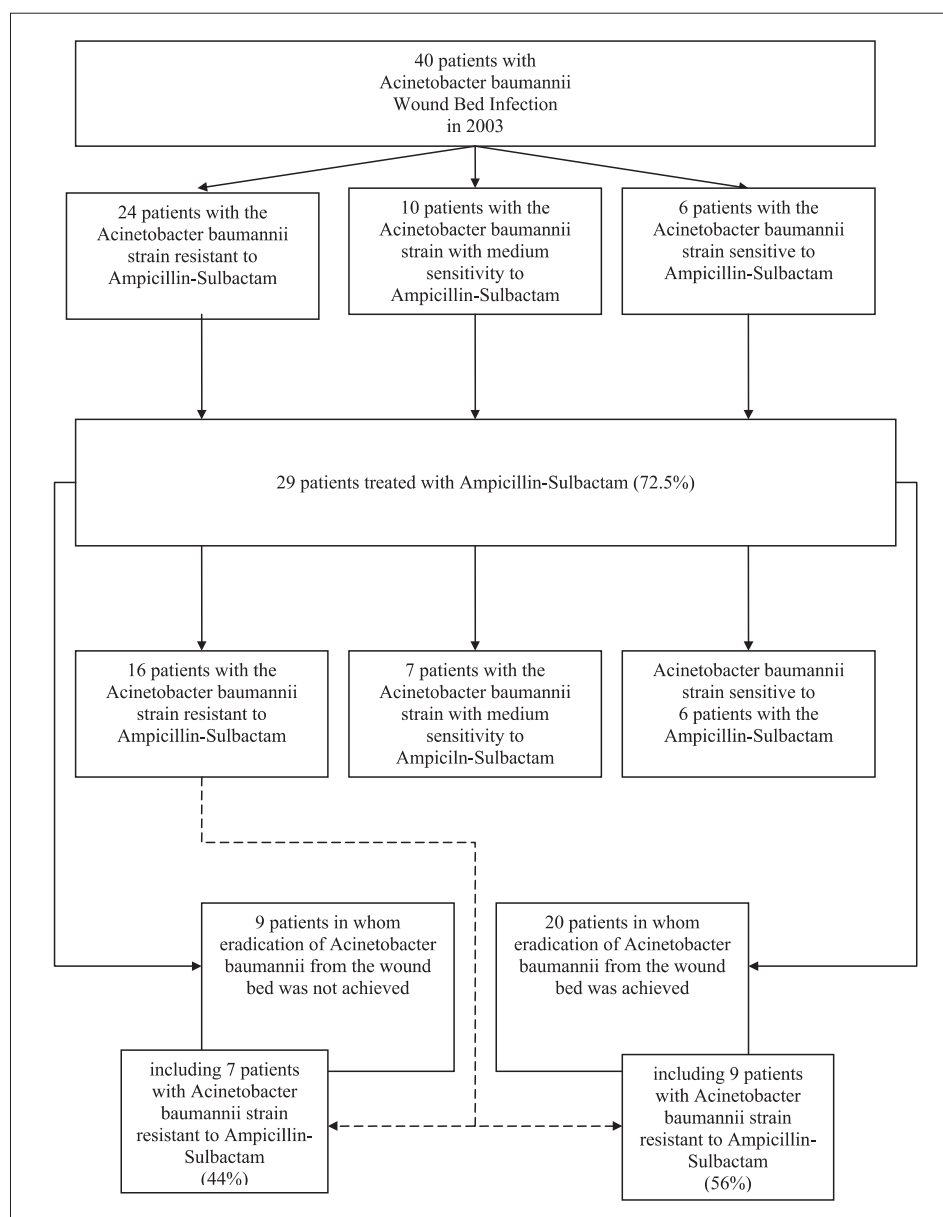
On the 6th of September, 2001, the Polish health authorities introduced a law on infectious diseases. A registration system regarding hospital infections was put in place. It is mandatory for all health care institutions to control methods of infection treatment, such as the use of antibiotics. Institutions must develop and implement infection control committees, working with specialized infection control teams, etc.

In close collaboration with the infection control team, the committee analyses the use of antibiotics, against the

background of frequently occurring infections, their etiology and the ever-changing sensitivity to antibiotics. These teams must jointly develop the strategies to limit an increase in resistance to antibiotics.

Correct identification of the AB species along with other “dangerous pathogens” with genotypical methods is necessary, to ensure awareness of the role of various genomic species of AB within the hospital epidemiology and the associated correct antibiotic treatment. Before starting treatment of AB bacteremia it is important to conduct a clinical examination of the patient in order to eliminate the possibility of pseudo - bacteremia and thereby avoiding unnecessary treatment [12]. Incorrect diagnostics may be a cause for treatment failures. The clinical symptoms of bacteremia caused by AB are not specific. Bacteremia caused by AB strains has a high mortality and is difficult to distinguish from other caus-

Table 8. Analysis of Burn Patients with a Wound Bed Infected by Strains of *Acinetobacter baumannii* in 2003



es of morbidity and mortality in burn patients [12] (Table 2).

In the past the AB strain was determined on the basis of increase in resistance. This way there was no certainty as to whether it was the same - or several clones. Now errors in diagnostics in our Centre are reduced as standards for collection of materials and microbiological analysis were introduced.

The reaction of AB to antibiotic treatment with AS may depend on factors, such as:

- Factors associated with the patient, i.e., age, condition of the immune system, trauma and its severity, level of plasma proteins, etc
- Factors associated with the microorganisms causing the infection, i.e. virulence

The more extensive the burn and the older the patient, the lower the albumin blood level may be. As a result the administered antibiotics may be less effective. Younger patients with less extensive burns and a higher albumin blood-level may respond better to the treatment. (Table 4, 5).

The microorganisms that caused the infection may also influence the efficacy AS therapy. AB has various properties influencing its virulence, such as the ability to create a biological membrane or biofilm. This is not only a method for linking the bacterial cell with the host cell, but may also influence the microorganism's virulence.

This biofilm creates resistance to the defense mechanisms of the host. It may disrupt phagocytosis, suppress antigens and/or breaks down the antibiotic and the antibodies.

The created biofilm may undergo fragmentation, together with the microorganisms present, it may be released into the blood circulation causing bacteremia, septic blockages and sources for secondary infection. With the disintegration of the cell, release of endotoxins occurs, which may lead to septic shock.

Based on the factors influencing the effectiveness of antibiotic treatment, we suggest that a biofilm was not formed in 9/29 patient group with the multi-resistant AB strain (Table 8). Due to their level of albumin, adequate

linkage with the antibiotic was created and full eradication of AB from the wound bed occurred, although AB was shown, in vitro, resistant to AS.

In 7/29 patients we noted that despite application of the full AS course, eradication of AB from their blood was not achieved. This may be caused by a reduced permeability of the cell membrane to AS, due to the formation of a biofilm and/or blood levels of proteins too low to bond with the antibiotic.

Despite the course set in, we still face a number of issues, such as what determined the success and failure of AB treatment with AS in the patients observed in our study?

The adequate treatment of patients that were suffering from a multi-microbial type of infection, such as gram-positive flora or non-oxygenated flora, as well as correct diagnosis and microbiological identification, remains difficult (Table 6, 7).

CONCLUSIONS

1. Correct diagnosis is mandatory to improve outcome of treatment. In order to determine whether an outbreak of AB is caused by the same clone of *Acinetobacter baumannii*, it is necessary to conduct genotype identification.
2. In treating difficult infections caused by *Acinetobacter baumannii*, combined treatment with several antibiotics should be considered to improve clinical outcome.
3. For wound bed infection caused by multi-resistant strains of AB, despite in vitro resistance, the application of Ampicillin with Sulbactam in a maximum daily dose, may be an effective treatment. It was noted to be an alternative to netilmicin.
4. In order to limit *Acinetobacter baumannii* outbreaks it is necessary to develop and implement standard procedures for diagnostics and treatment, such as an adequate antibiotic policy.

For the future other than pharmacological methods for treating infections are required such as vaccination and/or use of bacteriophages.

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